Supplementary Figure 1

A) FucT-IV / GAPDH mRNA copies (x10^-4) vs Days of co-culture

B) C2GlcAcT / GAPDH mRNA copies (x10^-3) vs Days of co-culture

C) CCR10 / GAPDH mRNA copies (x10^-4) vs Days of co-culture

D) α4 chain / GAPDH mRNA copies (x10^-4) vs Percent of Input Cells

E) αE chain / GAPDH mRNA copies (x10^-5) vs Percent of Input Cells

F) β1 chain / GAPDH mRNA copies (x10^-4) vs Percent of Input Cells

G) α4 chain (MFI) vs Days of co-culture

H) β7 chain (MFI) vs Days of co-culture

I) Chemotactic index vs Medium / CCL27

J) Chemotactic index vs Medium / CCL1

K) % of input cells vs Medium / CCL27

L) % of input cells vs Medium / CCL1
Supplementary Figure 1. mRNA expression, integrin chain expression, and chemotactic responsiveness of CD8 T cells activated with PLN-DC or PP-DC.

Naïve CD8 T cells were cultured with peptide-pulsed PLN-DC or PP-DC. The resulting effector CD8 T cells were analyzed on consecutive days for expression of mRNAs for A) FucT-IV (n=3), B) C2GlcNAcT-I, C) CCR10 (n=3), and after 5-6 days of coculture for D) α4, β7, αE and β1 integrin chain (results show the average of 1 experiment performed in triplicate). E) Surface expression level of α4, β7, αE and β1 integrin chain. To compare CD8 T cells activated with PLN-DC vs. CD8 T cells activated with PP-DC, a two-tailed paired Student’s t test was used (**p<0.01, *p<0.05, n=5-10). F,G) Chemotaxis experiments were performed after 4-5 days of coculture using F) 100 nM of the CCR10 agonist CCL27 (n=5) or G) the CCR8 agonist CCL1 (n=1 experiment performed in duplicate). Data are expressed as mean ± SE.

Primers used for real-time PCR:

FucT-IV
Forward:TCTAGCCTTTTGAGAACCTCAG; Reverse:TCATAGTTGGCACGATCTGG

C2GlcNAcT-I
Forward: CAGGAGTCAGAGCCTCAACAGA; Reverse:TGCCAGTTTATCAGCGGGAC

CCR10 (A and B isoforms)
Forward:CTGTTGTCGCTTCTCAGA; Reverse:TTTCACAGTCTGCGTGGAGG
Supplementary Figure 2. Effect of sorted DC subsets in the acquisition of selectin ligands and α4β7 on CD8 T cells.

DC from PP, PLN and spleen were isolated by negatively selection and then subdivided by FACS sorting into CD11c^{Low}B220^{+} (‘plasmacytoid’ DC) and CD11c^{High}B220^{Neg} DC. The latter were further sorted into CD8α^{High}CD11b^{Low} (‘lymphoid’ DC), CD8α^{Low}CD11b^{High} (‘myeloid’ DC) and CD8α^{Low}CD11b^{Low} subsets. Each DC subpopulation was pulsed with peptide Ag and cocultured with naïve CD8 T cells. The resulting effector CD8 T cells were analyzed after 5 days for the expression of A) E-Lig, B) P-Lig, C) α4β7, or D) LFA-1. Data are expressed as mean ± range (n=2).
Supplementary Figure 3. Correlation between the expression of the homing molecules $\alpha_4\beta_7$, CCR9 and ligands for E- and P-selectin on endogenous effector/memory T cells used for FACS sorting.

CD8+ T cells were isolated by negative selection from pooled cells from spleen, PLN, MLN and PP from 4-6 month old C57Bl/6 mice. For the analysis, cells were gated on viable (FSC/SSC), CD8+CD44$^{\text{High}}$ effector/memory T cells. Dotted line was based on appropriate isotype controls.